

Dietary nitrate's effects on exercise performance in heart failure with reduced ejection fraction (HFrEF)

Vinaya Mulkareddy MD^a,

Susan B. Racette PhD^{a,b},

Andrew R. Coggan PhD^c,

Linda R. Peterson MD^d

Institutional affiliations: ^aThe Department of Medicine, 4960 Children's Place, Campus Box 8066, St. Louis, MO 63110, USA; ^bProgram in Physical Therapy at Washington University School of Medicine, Campus Box 8502, 4444 Forest Park Ave. St. Louis, Missouri, 63108-2212, USA; ^cDepartments of Kinesiology and Cellular and Integrative Physiology at Indiana University-Purdue University at Indianapolis, 901 West New York Street, Indianapolis, IN, 46202 USA, and ^dDepartment of Medicine, Cardiovascular Division at Washington University School of Medicine, St. Louis, Missouri, USA. The e-mail addresses of the authors are: vmulkare@gmail.com, racettes@wustl.edu, acoggan@iupui.edu, and lpeterso@wustl.edu, respectively.

Corresponding author:

Linda R. Peterson, MD
Washington University School of Medicine
Campus Box 8086,
660 S. Euclid Ave.
Saint Louis, Missouri, USA 63110
e-mail lpeterso@wustl.edu
Facsimile 314-362-9982

This is the author's manuscript of the article published in final edited form as:

Mulkareddy, V., Racette, S. B., Coggan, A. R., & Peterson, L. R. (2018). Dietary nitrate's effects on exercise performance in heart failure with reduced ejection fraction (HFrEF). *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*. <https://doi.org/10.1016/j.bbadis.2018.09.026>

Abstract:

Heart failure with reduced ejection fraction (HFrEF) is a deadly and disabling disease. A key derangement contributing to impaired exercise performance in HFrEF is decreased nitric oxide (NO) bioavailability. Scientists recently discovered the inorganic nitrate pathway for increasing NO. This has advantages over organic nitrates and NO synthase production of NO. Small studies using beetroot juice as a source of inorganic nitrate demonstrate its power to improve exercise performance in HFrEF. A larger-scale trial is now underway to determine if inorganic nitrate may be a new arrow for physicians' quiver of HFrEF treatments.

Keywords: Nitric oxide, inorganic nitrate, exercise performance, heart failure, muscle power, peak oxygen consumption

Abbreviations: HF = heart failure, HFrEF = heart failure with reduced ejection fraction, cGMP = cyclic guanine monophosphate, sGC = soluble guanylyl cyclase, NO = nitric oxide, NO_3^- = nitrate, NO_2^- = nitrite, NOS = nitric oxide synthase, ROS = reactive oxygen species, ALDH-2 = aldehyde dehydrogenase, PDE5 = phosphodiesterase 5, PKG = cGMP-dependent protein kinase, SERCA = sarcoendoplasmic reticulum calcium transport ATPase.

Introduction.

Heart failure (HF), specifically heart failure with reduced ejection fraction (HFrEF), is a major public health problem. HFrEF is not only a deadly but also a disabling disease. This disability is thought to relate, at least in part, to impaired nitric oxide (NO) bioavailability. Recent discoveries regarding the inorganic nitrate pathway as a novel source of NO for the body have led to a burgeoning field of research leveraging this pathway for the treatment of diseases including HFrEF. This pathway has particular advantages over other mechanisms for the production of NO. Moreover, most preliminary studies show that inorganic nitrate can increase both muscle power and aerobic exercise performance, which should improve patients' ability to perform their activities of daily living. Continued research is ongoing to determine if inorganic nitrate may be a novel, effective treatment for HFrEF.

Significance of heart failure with reduced ejection fraction (HFrEF).

Heart failure is a public health problem of epidemic proportion. It affects ~6 million people in the U.S. and ~23 million worldwide [1, 2]. Unfortunately, these numbers are increasing. Adults over age 40 y now have a 1 in 5 chance of developing HF in their lifetime. Moreover, HF costs ~\$39.1 billion annually in the U.S. alone. The human cost is great as well: 40% of patients die within 1 y of diagnosis and 5 y survival is only 35% [1]. In addition, those who live with HF often endure significant disability and decreased quality of life. Given the costly and disabling nature of HF and its tremendous impact on quality of life, any improvement in exercise performance would be of enormous benefit to patients with this disease.

HFrEF is a particularly disabling disease in part because it impairs patients' capacity to perform aerobic exercise and increases ventilatory effort, resulting in dyspnea. Indeed, the main

classification systems used to evaluate HFrEF stratify the severity of disease based on the physical limitations it imposes. For example, as the New York Heart Association class of HF increases (from I to IV), the limitations in aerobic exercise increase. Demonstrating impaired aerobic exercise tolerance in HF is an important component of declaring a patient “disabled” due to HF. Moreover, impaired aerobic capacity during exercise is one of the key tests used to evaluate HFrEF patients for cardiac transplantation and to predict mortality [3, 4]. The lower the peak oxygen consumption ($\dot{V}O_{2peak}$) in HF patients, the worse the prognosis and the worse the disability.

Decreased skeletal muscle *power* (speed x force of contraction) also impairs quality of life and predicts increased mortality. Most typical daily activities require muscle power, as opposed to aerobic capacity. Getting out of bed, rising from a chair, opening a jar, lifting groceries, picking up a child, and climbing stairs are all activities that *require power*. Although it is well-known that HFrEF impairs aerobic capacity, it is less widely appreciated that HFrEF also decreases muscle strength and power [5, 6]. HFrEF patients are less powerful even when compared to equally-sedentary but healthy individuals with comparable limb muscle mass, indicating that the muscular deficits in HFrEF are not simply due to physical inactivity or muscle atrophy [6]. Instead, recent studies have demonstrated that the muscle dysfunction found in HF patients is characterised by derangements at the molecular level [7-10]. In HF, decreased muscle power also predicts increased mortality [11]. In one study of more than 100 HF patients, decreased skeletal muscle power was a more powerful predictor of mortality than $\dot{V}O_{2peak}$ (Figure 1) [12]. Even in young healthy adults, lower muscle power predicts increased mortality. In a study of more than 1 million young men, all-cause mortality was a striking *122.3 per 100,000 person-years in the weakest versus 5.6 per 100,000 person-years in the strongest men* [13]. Clearly, muscle power is an extremely important target for treatment in HF, yet is one that is not presently addressed by any standard medications or therapies.

NO deficiency — a key derangement in HFrEF.

Numerous factors account for the decline in exercise performance in HFrEF patients. These include, but are not limited to, increased skeletal muscle breakdown, increased oxidative stress, inflammation, and hypoperfusion. An excellent review of the many mechanisms that affect skeletal muscle function in heart failure is provided by Schulze and Toth in, Heart Failure, A Companion to Braunwald's Heart Disease [14]. It is beyond the scope of this mini-review to detail all of these factors; instead, we will focus on one key molecular factor contributing to these derangements — low NO bioavailability [15]. The evidence for decreased NO bioavailability is manifold. Breath NO levels are lower in patients with HFrEF compared with healthy individuals [16]. Decreased plasma levels of nitrosothiols and cyclic guanosine monophosphate (cGMP) — a key mediator of NO effects — also indicate low NO bioavailability in HFrEF [17]. NO stimulates guanyl cyclase (sGC) to increase cGMP production, which has opposite effects in smooth and skeletal muscle. In smooth muscle (e.g., in the arterial wall), NO causes vasodilation via stimulation of sGC and hence increased cGMP. In skeletal muscle, NO stimulation of sGC and upregulation of cGMP increases force of contraction [18] and the capacity for mitochondrial fatty acid oxidation [19]. To be sure, there are other pathways by which NO affects muscle. It is evident that NO deficiency could cause a decrease in cGMP and consequent impairments in vasodilation and aerobic exercise capacity, as well as decreased muscle power. Indeed, patients with HFrEF have impaired endothelial function, as was demonstrated by human cardiac catheterization studies in the 1990s [17]. Importantly, this impaired endothelial function in HFrEF is independently associated with an increased incidence of HF hospitalization, cardiac transplantation, and death [20].

The mechanisms by which NO bioavailability is reduced in HFrEF include both decreased production and enhanced degradation of NO [17]. In HFrEF, the activity of the endothelial isoform of NO synthase, eNOS, is decreased [17], while the levels of reactive oxygen species (ROS) that degrade NO are increased [21]. ROS levels are higher, at least in part, due to decreased antioxidant defenses [17]. Moreover, this increased oxidative stress in the left ventricle is correlated with the severity of HFrEF [22]. Studies in animal models further support the idea that decreased NO bioavailability is pathophysiologically linked to HF, rather than simply associative. These murine studies demonstrate a protective effect of enhanced production of NO (via eNOS overexpression) against HF development. Consistent with this premise that eNOS is beneficial, animal models that are deficient in eNOS are more susceptible to HF development, left ventricular hypertrophy, and hypertension [17]. Thus, increasing NO bioavailability in HFrEF patients is an attractive target for the amelioration of HFrEF symptoms.

Nitric oxide (NO) production: the main metabolic pathways.

There are 3 main pathways for increasing NO production, as shown in Figure 2. The pharmacologic, organic nitrate pathway has the longest history in Western medicine, with drugs such as nitroglycerin (glyceryl trinitrate) used for over 150 years to ameliorate angina and HFrEF symptoms due to its vasodilatory effects. However, it wasn't until almost 100 years later that scientists discovered that NO was the primary molecule responsible for the vasodilatory effects. Nitroglycerin generates NO through mitochondrial aldehyde dehydrogenase (ALDH-2) [23] by generating the intermediary products nitrate (NO_3^-) and 1,2- glyceryl dinitrate; the NO_3^- in the mitochondria is subsequently reduced to NO and/or converted to S-nitrosothiol. An important disadvantage of this pathway is that prolonged organic NO_3^- treatment often induces tolerance (i.e., impaired vasodilation response to nitroglycerin treatment) and cross-tolerance (i.e., impaired endothelium-dependent vasodilation), with oxidative stress playing an important

role. Data suggest that inactivation of a key enzyme in the processing of nitroglycerin — mitochondrial ALDH-2 — by reactive oxygen species (ROS) is central to tolerance and cross-tolerance [24]. Chronic nitroglycerin treatment can also lead to supersensitivity to vasoconstrictive molecules through chronic activation of protein kinase C [23]. Different organic nitrates have varying propensities for inducing tolerance [25] and their use may be limited by side effects such as severe headaches, hypotension, and rebound vasoconstriction, a phenomenon that occurs following nitrate withdrawal and which may be attributable to coronary vasoconstriction.

A second pathway by which the body can synthesize NO is the endogenous nitric oxide synthase (NOS) pathway (Figure 2) discovered by Furchgott, Ignarro, and Murad, who were awarded the 1998 Nobel Prize in Physiology or Medicine [26] *“for their discoveries concerning nitric oxide as a signaling molecule in the cardiovascular system”* (Nobelprize.org). In this endogenous pathway, NO is synthesized enzymatically from the conditionally essential amino acid L-arginine, oxygen, and NADPH by three NOS isoforms: neuronal NOS (nNOS or NOS1), inducible NOS (iNOS or NOS2), and endothelial NOS (eNOS or NOS3). nNOS and eNOS are calcium dependent. In addition to NO, L-citrulline is produced and can be recycled to generate *de novo* arginine. Co-factors necessary in this process include tetrahydrobiopterin (BH4), flavin adenine dinucleotide (FAD), flavin mononucleotide (FMN), and heme. There are limitations to this pathway, however. First, it requires oxygen. Thus, NO production by NOS enzymes decreases with ischemic duration, as shown by Giraldez et al. [27]. Second, NOS activity diminishes as pH decreases, with a marked lowering at pH <7.0. Not only do abnormal/diseased tissues have a pH in this range, but also healthy skeletal muscle that is engaged in vigorous exercise experiences pH decreases into this range [28]. Interestingly, the third NO production pathway works best in ischemic and/or acidic conditions, and thus is a perfect complement to the NOS pathway.

The third NO metabolic pathway is the dietary or inorganic nitrate pathway (also referred to as the enterosalivary pathway). Ironically, though this pathway likely has existed for millennia, it wasn't discovered until ~1994. There is a suggestion, however, that the effects of this dietary nitrate pathway were known in ancient China, where it was noticed that KNO_3 was useful for "treating symptoms...such as acute heart pains" [29]. In this 'enterosalivary' pathway, NO is produced through sequential reduction of *inorganic* dietary nitrate [29] independently of NOS, beginning with inorganic NO_3^- from dietary sources (e.g., beets and dark green leafy vegetables). Reduction to nitrite (NO_2^-) is facilitated by bacterial nitrate oxidoreductases from commensal bacteria in the oral cavity. The very low pH of the stomach also favors the chemical reduction of NO_3^- to NO_2^- , as well as the reduction of NO_2^- to NO. The intestine then absorbs the anions, which are taken up by the circulation [29]. Once in the circulation, the skeletal muscle may take up nitrate and serve as an important 'reservoir' of nitrate (Figure 2) [30]. Further acidic reduction of NO_2^- to NO occurs in tissues, catalyzed by deoxy-hemoglobin, deoxy-myoglobin, xanthine oxidoreductase, or other nitrate reductases. Heavily exercising skeletal muscle is also a prominent tissue for the final conversion to NO because the metabolic conditions are appropriate (i.e., low pH and low pO_2) [28]. Of note, this pathway can also move in reverse with decreasing acidity and abundant oxygen (e.g., nitrite can be made from NO). Recent data from Omar et al. show that this hypoxia-driven reduction of NO_2^- to NO can occur in *small resistance arterioles*, thus facilitating tissue perfusion [31]. Thus, this pathway is especially beneficial in hypoxia and ischemia. Although an estimated 75% of circulating nitrate is excreted by the kidneys or exhaled as NO, there is active re-uptake of approximately 25% of circulating nitrate by the salivary glands [32], which then secrete NO_3^- , thus re-starting the enterosalivary cycle of nitrate reduction in the oral cavity. The fact that the body actively recycles nitrate in this pathway underscores its biological importance. Indeed, estimates suggest that a significant fraction of the body's NO is derived from this exogenous pathway. It is also interesting to note

that Omar et al. [31] showed that supraphysiologic/near-physiologic levels of inorganic NO_2^- can be reduced to NO in normoxic – but not hypoxic or hyperoxic – conditions in *conduit* arteries. In short, the inorganic NO_3^- pathway for NO generation is complex, likely has been operative in mankind for millennia, and functions in conditions not conducive to NO production by NOS. There are also several other specific advantages of this inorganic nitrate pathway as a source for NO generation over the other two pathways.

Advantages of the inorganic nitrate pathway as a source of NO for the cardiovascular system.

The exogenous inorganic nitrate pathway for generation of NO has distinct advantages over the NOS pathway and over organic nitrate drugs (see Table 1). As mentioned above, the inorganic nitrate pathway functions best in ischemic and acidic conditions, particularly at the arteriolar level [31], thus facilitating NO-related effects such as vasodilation and skeletal muscle contraction, particularly in stressed or diseased tissues. The NOS pathway, by contrast, is O_2 -dependent and therefore does not function well under these conditions. The inorganic pathway also has a slower onset of action than some organic nitrates, which may contribute to fewer reports of hypotension, flushing, and severe headaches [33-36]. There are also reports that inorganic nitrate treatment lessens mitochondrial oxidative stress [37], whereas organic nitrates, such as nitroglycerin, increase it [23]. As mentioned above, this increase in oxidative stress and other mechanisms cause tolerance with continuous use of organic nitrates such as nitroglycerin. Importantly, there are data from studies of continuous infusion of inorganic nitrite in nonhuman primates [38] and in humans [39] showing that there is no tolerance to inorganic nitrite. A recent study by Andrew Murray's group demonstrated another benefit of the inorganic pathway: dietary inorganic nitrate increases arginine availability for the NOS pathway by suppressing cardiac arginase expression and increasing tissue L-arginine in both normoxic and hypoxic

conditions [37]. Recent data from Chirinos et al. in patients with HF with preserved ejection fraction also show that inorganic nitrate reduces the left ventricular load late in systole, which is caused by reflected waves from the arterial tree. Inorganic nitrate administration shifts the reflected wave later into diastole, thereby improving coronary perfusion pressure [40]. Table 1 also includes advantages of inorganic nitrate therapy over sildenafil and other phosphodiesterase 5 (PDE5) inhibitors; these drugs also increase cGMP, induce vasodilation, and have other cGMP-mediated effects, but do not increase NO. Thus, there are several advantages of the inorganic nitrate pathway over other pathways for NO production, and eventually, cGMP amplification. These advantages make inorganic nitrate an attractive potential therapeutic agent for increasing NO, especially in patients who have reduced NO bioavailability – such as those with HFrEF.

Inorganic nitrate effects on exercise performance in patients with HFrEF.

Preliminary data show that inorganic nitrate improves muscle power and NO bioavailability. In one small, double-blind, placebo-controlled study, 11.2 mmol of inorganic nitrate, given in the form of concentrated beetroot juice, increased quadriceps muscle power within two hours *after a single dose* [34]. Peak muscle power was increased by ~11% at the highest movement velocity tested. The inorganic nitrate consumption was accompanied by a ~20 fold increase in plasma nitrate levels, as well as a 35-50% increase in breath NO levels [34]. Calculated maximal velocity of contraction and maximal power were also significantly increased after ingestion of dietary inorganic nitrate (13 and 12%, respectively) [34]. The improvement in muscle power seen after a single dose of inorganic nitrate is comparable to that which would be expected to result from ~2 to 3 months of resistance exercise training [34]. Based on a study by Toth et al. [6], in which patients with HFrEF were shown to have less powerful muscle function (even after correcting for muscle mass and other variables), the improvements observed after inorganic

nitrate treatment would have erased ~1/3 of the HF-related deficit in muscle power [34]. This improvement in muscle power after inorganic nitrate therapy contrasts with the lack of an effect that standard HF therapies (i.e., beta-adrenergic antagonists, angiotensin-converting enzyme inhibitors, angiotensin receptor blockers, and aldosterone antagonists) have on muscle power [34]. It is unclear whether inorganic nitrate can also increase myocardial contractile power, as it does in skeletal muscle. Nevertheless, an improvement in skeletal muscle function alone should help patients with HFrEF perform many of their activities of daily living because many activities rely on muscle power.

Results from studies on the effect of inorganic nitrate on aerobic exercise capacity in patients with HFrEF are slightly more mixed. Coggan et al. showed in a small, double-blind, placebo-controlled study that dietary inorganic nitrate markedly increased plasma nitrate, plasma nitrite, and breath NO levels (by $1469 \pm 245\%$, $105 \pm 34\%$, and $60 \pm 18\%$, respectively) [41]. Simultaneous with this increase in NO bioavailability, $\dot{V}O_{2peak}$ increased by $8 \pm 2\%$ and time to fatigue increased by $7 \pm 3\%$. Exercise efficiency, however, did not change [41]. Although this increase in $\dot{V}O_{2peak}$ is small, based on outcomes studies, this change in $\dot{V}O_{2peak}$ should translate into a ~10% decrease in the annual risk of death or transplant [41, 42].

The improvement in $\dot{V}O_{2peak}$ seen in the study by Coggan et al. correlates well with results of a study by Kerly et al., in which a single dose of inorganic nitrate enhanced incremental shuttle walk test performance in patients with HFrEF by ~18% as compared with placebo [43]. Of note, these positive effects on aerobic performance also correlate with a study of inorganic nitrate's effects in HF patients with *preserved* ejection fraction [36]. A recent study by Hirai et al. in male patients, most of whom had ischemic cardiomyopathy, however, did not show an improvement in $\dot{V}O_{2peak}$ after repeated dosing of inorganic nitrate [44]. Differences in HFrEF etiology, other patient characteristics, or nitrate dosing regimens between studies may account for the differences in $\dot{V}O_{2peak}$ results. Although the source of inorganic nitrates was

beetroot juice in both the Coggan [41] and Hirai [44] studies, the inorganic nitrate dose that was ingested 2 hours before exercise testing was 11.2 mmol in the Coggan study and 6.45 mmol in the Hirai study. The difference in $\dot{V}O_{2peak}$ results does not appear to be due to differences in HFrEF severity or ejection fraction.

Mechanisms of action of NO relating to improved exercise performance in HFrEF.

There are several NO-mediated effects on muscle contractile function [18]. Because the net effect of inorganic nitrate ingestion in patients with HFrEF is an improvement in muscle contractile function, the NO-related pathways that increase muscle contractile function must 'win' over the NO-related pathways that would decrease muscle function. As mentioned above, NO increases sGC activity, thereby increasing cGMP. This leads to an increase in the speed of muscle shortening, and hence an increase in power [18]. Increased NO availability can also result in nitrosylation of the ryanodine receptor on the sarcoplasmic reticulum, which opens this channel [45]. This may result in an increase in calcium release by the sarcoplasmic reticulum and subsequent enhancement of both maximal muscle contraction velocity and power. Other NO-related effects (e.g., nitrosylation of troponin I [46] or myosin [47] that would be expected to have a negative effect on muscle contractile function appear to be weaker than these positive effects. The sum of these pleiotropic NO-related effects in vivo in patients with HFrEF appears to result in increased muscle power, which should translate into improved functional capacity.

Several NO-mediated mechanisms also likely contribute to the general improvement in aerobic capacity after inorganic nitrate administration. Enhanced efficiency of mitochondrial oxidative phosphorylation is one mechanism implicated in the reduced oxygen costs of exercise [48-50]. The increase in $\dot{V}O_{2peak}$ described by Coggan et al. in patients with HFrEF likely resulted from an increase in cardiac output and/or arterio-venous O_2 difference at peak exercise [41]. In that study, peak diastolic blood pressure trended lower and heart rate trended higher

after inorganic nitrate ingestion, suggesting a higher cardiac output accompanying lower peripheral vascular resistance [41]. Since NO is known to be the original “endothelium relaxing factor”, as coined by Furchgott, it is reasonable to conclude that the arteriolar vasodilating effects of NO may contribute to an improvement in cardiac output and, hence, $\dot{V}O_{2peak}$. Studies of infusions of inorganic nitrite in patients with HFrEF corroborate this, as they show that increased forearm blood flow increases in the infused arm [39]. There are also data from animal models of NOS manipulation and in vitro studies that indicate that NO from NOS may also have direct, positive cardiac lusitropic, inotropic, and/or chronotropic effects distinct from responses to vasodilation. These effects were detailed in a review by Massion et al. [51]. Data from patients with heart failure with *preserved* ejection fraction show that inorganic nitrate, in contrast to organic nitrates, can also lessen late systolic pulsatile left ventricular load. Whether one or all of these direct cardiac and pulsatile load effects are active in patients with HFrEF remains unclear.

Summary – Conclusions

NO is a powerful radical with pleiotropic effects. It is of particular therapeutic interest in the treatment of HFrEF because NO bioavailability is low in HFrEF patients and is thought to be a key mediator of HF pathophysiology. There are three main pathways that can be leveraged to increase NO bioavailability. The last to be discovered, the inorganic nitrate pathway, has several advantages over the endogenous and organic pharmacologic pathways, as well as over PDE5 inhibitors. Inorganic nitrates have been shown in most small clinical studies to improve exercise performance – both muscle power and aerobic exercise performance. A larger study, the INIX-HF trial (NCT02797184), is now underway to perform the studies necessary and sufficient to set up a multicenter study of the effectiveness of inorganic nitrate for the improvement of exercise performance and quality of life in patients with HFrEF.

Conflicts of interest: The authors have no conflicts of interest to declare.

Acknowledgements: This work was supported by the National Institutes of Health (NIH) [1R34HL138253-01]; the Barnes-Jewish Hospital Foundation; the Mentors in Medicine and C-STARS programs at Washington University School of Medicine.

Figure Legends

Figure 1. Muscle power as a predictor of survival in patients with HFrEF [12]. Kaplan Meier lifetime analysis of survival stratified by peak torque index of the knee flexor muscles at a cut-off point of 68 N-m x 100 per kg body weight.

Figure 2. The main pathways for nitric oxide (NO) production. The dietary pathway (starting at upper left) utilizes *inorganic* nitrates and is facilitated by lower pO_2 and pH. Ingestion of nitrate-containing foods, especially high-nitrate foods such as beetroot are the source of the inorganic nitrate (NO_3^-) start this pathway. NO_3^- is reduced to nitrite (NO_2^-) by reductases or acidic conditions and facilitated by oxyheme proteins. Then NO_2^- is reduced to NO under the appropriate conditions. Importantly, skeletal muscle can serve as a ‘reservoir’ for nitrate [30]. As shown by the dashed arrows, this pathway can also ‘run in reverse’ with NO being used to create NO_2^- and then NO_3^- given the appropriate conditions.

*Note: NO_3^- can be taken up from the circulation into the salivary glands and go through this reduction pathway again in what is known as ‘the enterosalivary pathway.’

The endogenous pathway (lower left) uses NO synthase and oxygen to create citrulline and NO. An abbreviated depiction of the organic nitrate pathway (lower right) shows the production of NO derived from pharmacologic sources, such as nitroglycerin.

ALDH-2 = aldehyde dehydrogenase, P450 = cytochrome P450.

References

- [1] A.S. Go, D. Mozaffarian, V.L. Roger, E.J. Benjamin, J.D. Berry, M.J. Blaha, S. Dai, E.S. Ford, C.S. Fox, S. Franco, H.J. Fullerton, C. Gillespie, S.M. Hailpern, J.A. Heit, V.J. Howard, M.D. Huffman, S.E. Judd, B.M. Kissela, S.J. Kittner, D.T. Lackland, J.H. Lichtman, L.D. Lisabeth, R.H. Mackey, D.J. Magid, G.M. Marcus, A. Marelli, D.B. Matchar, D.K. McGuire, E.R. Mohler, C.S. Moy, M.E. Mussolino, R.W. Neumar, G. Nichol, D.K. Pandey, N.P. Paynter, M.J. Reeves, P.D. Sorlie, J. Stein, A. Towfighi, T.N. Turan, S.S. Virani, N.D. Wong, D. Woo, M.B. Turner, Heart disease and stroke statistics--2014 update: a report from the American Heart Association., *Circulation* 129 (2014) e28-e292.
- [2] J.J. McMurray, M.C. Petrie, D.R. Murdoch, A.P. Davie, Clinical epidemiology of heart failure: public and private health burden., *Eur Heart J* 19 Suppl P (1998) P9-16.
- [3] D.M. Mancini, H. Eisen, W. Kussmaul, R. Mull, L.H.J. Edmunds, J.R. Wilson, Value of peak exercise oxygen consumption for optimal timing of cardiac transplantation in ambulatory patients with heart failure., *Circulation* 83 (1991) 778-786.
- [4] L.R. Peterson, P. Herrero, K.B. Schechtman, S.B. Racette, A.D. Waggoner, Z. Kisrieva-Ware, C. Dence, S. Klein, J. Marsala, T. Meyer, R.J. Gropler, Effect of obesity and insulin resistance on myocardial substrate metabolism and efficiency in young women., *Circulation* 109 (2004) 2191-2196.
- [5] D. Harrington, S.D. Anker, T.P. Chua, K.M. Webb-Peploe, P.P. Ponikowski, P.A. Poole-Wilson, A.J. Coats, Skeletal muscle function and its relation to exercise tolerance in chronic heart failure., *J Am Coll Cardiol* 30 (1997) 1758-1764.
- [6] M.J. Toth, A.O. Shaw, M.S. Miller, P. VanBuren, M.M. LeWinter, D.W. Maughan, P.A. Ades, Reduced knee extensor function in heart failure is not explained by inactivity., *Int J Cardiol* 143 (2010) 276-282.

- [7] M.S. Miller, P. Vanburen, M.M. Lewinter, S.H. Lecker, D.E. Selby, B.M. Palmer, D.W. Maughan, P.A. Ades, M.J. Toth, Mechanisms underlying skeletal muscle weakness in human heart failure: alterations in single fiber myosin protein content and function., *Circ Heart Fail* 2 (2009) 700-706.
- [8] M.P. Godard, S.A. Whitman, Y.H. Song, P. Delafontaine, Skeletal muscle molecular alterations precede whole-muscle dysfunction in NYHA Class II heart failure patients., *Clin Interv Aging* 7 (2012) 489-497.
- [9] H.R. Middlekauff, C. Vigna, M.A. Verity, G.C. Fonarow, T.B. Horwich, M.A. Hamilton, P. Shieh, A.R. Tupling, Abnormalities of calcium handling proteins in skeletal muscle mirror those of the heart in humans with heart failure: a shared mechanism?, *J Card Fail* 18 (2012) 724-733.
- [10] E. Rullman, D.C. Andersson, M. Melin, S. Reiken, D.M. Mancini, A.R. Marks, L.H. Lund, T. Gustafsson, Modifications of skeletal muscle ryanodine receptor type 1 and exercise intolerance in heart failure., *J Heart Lung Transplant* 32 (2013) 925-929.
- [11] E.G. Artero, D.C. Lee, C.J. Lavie, V. Espana-Romero, X. Sui, T.S. Church, S.N. Blair, Effects of muscular strength on cardiovascular risk factors and prognosis., *J Cardiopulm Rehabil Prev* 32 (2012) 351-358.
- [12] M. Hulsmann, M. Quittan, R. Berger, R. Crevenna, C. Springer, M. Nuhr, D. Mortl, P. Moser, R. Pacher, Muscle strength as a predictor of long-term survival in severe congestive heart failure., *Muscle strength as a predictor of long-term survival in severe congestive heart failure*. 6 (2004) 101-107.
- [13] F.B. Ortega, K. Silventoinen, P. Tynelius, F. Rasmussen, Muscular strength in male adolescents and premature death: cohort study of one million participants., *BMJ* 345 (2012) e7279.
- [14] D.L. Mann, G.M. Felker, *Heart Failure*, (2016)

- [15] W.J. Paulus, C. Tschope, A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation., *J Am Coll Cardiol* 62 (2013) 263-271.
- [16] H. Adachi, S. Oshima, S. Sakurai, T. Toyama, H. Hoshizaki, K. Taniguchi, H. Ito, Nitric oxide exhalation correlates with ventilatory response to exercise in patients with heart disease., *Eur J Heart Fail* 5 (2003) 639-643.
- [17] S. Bhushan, K. Kondo, D.J. Polhemus, H. Otsuka, C.K. Nicholson, Y.X. Tao, H. Huang, V.V. Georgiopoulou, T. Murohara, J.W. Calvert, J. Butler, D.J. Lefer, Nitrite therapy improves left ventricular function during heart failure via restoration of nitric oxide-mediated cytoprotective signaling., *Circ Res* 114 (2014) 1281-1291.
- [18] G. Marechal, P. Gailly, Effects of nitric oxide on the contraction of skeletal muscle., *Cell Mol Life Sci* 55 (1999) 1088-1102.
- [19] T. Ashmore, L.D. Roberts, A.J. Morash, A.O. Kotwica, J. Finnerty, J.A. West, S.A. Murfitt, B.O. Fernandez, C. Branco, A.S. Cowburn, K. Clarke, R.S. Johnson, M. Feelisch, J.L. Griffin, A.J. Murray, Nitrate enhances skeletal muscle fatty acid oxidation via a nitric oxide-cGMP-PPAR-mediated mechanism., *BMC Biol* 13 (2015) 110.
- [20] D. Fischer, S. Rossa, U. Landmesser, S. Spiekermann, N. Engberding, B. Hornig, H. Drexler, Endothelial dysfunction in patients with chronic heart failure is independently associated with increased incidence of hospitalization, cardiac transplantation, or death., *Eur Heart J* 26 (2005) 65-69.
- [21] H. Cai, D.G. Harrison, Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress., *Circ Res* 87 (2000) 840-844.
- [22] C. Maack, T. Kartes, H. Kilter, H.J. Schäfers, G. Nickenig, M. Böhm, U. Laufs, Oxygen free radical release in human failing myocardium is associated with increased activity of rac1-GTPase and represents a target for statin treatment., *Circulation* 108 (2003) 1567-1574.

- [23] T. Münzel, A. Daiber, A. Mülsch, Explaining the phenomenon of nitrate tolerance., *Circ Res* 97 (2005) 618-628.
- [24] K. Sydow, A. Daiber, M. Oelze, Z. Chen, M. August, M. Wendt, V. Ullrich, A. Mülsch, E. Schulz, J.F. Keaney, J.S. Stamler, T. Münzel, Central role of mitochondrial aldehyde dehydrogenase and reactive oxygen species in nitroglycerin tolerance and cross-tolerance., *J Clin Invest* 113 (2004) 482-489.
- [25] T. Munzel, S. Steven, A. Daiber, Organic nitrates: Update on mechanisms underlying vasodilation, tolerance and endothelial dysfunction., *Vascul Pharmacol* 63 (2014) 105-113.
- [26] R. SoRelle, Nobel prize awarded to scientists for nitric oxide discoveries., *Circulation* 98 (1998) 2365-2366.
- [27] R.R. Giraldez, A. Panda, Y. Xia, S.P. Sanders, J.L. Zweier, Decreased nitric-oxide synthase activity causes impaired endothelium-dependent relaxation in the postischemic heart., *J Biol Chem* 272 (1997) 21420-21426.
- [28] A.R. Coggan, A.M. Abduljalil, S.C. Swanson, M.S. Earle, J.W. Farris, L.A. Mendenhall, P.M. Robitaille, Muscle metabolism during exercise in young and older untrained and endurance-trained men., *J Appl Physiol* 75 (1993) 2125-2133.
- [29] J.O. Lundberg, E. Weitzberg, M.T. Gladwin, The nitrate-nitrite-nitric oxide pathway in physiology and therapeutics., *Nat Rev Drug Discov* 7 (2008) 156-167.
- [30] B. Piknova, J.W. Park, K.M. Swanson, S. Dey, C.T. Noguchi, A.N. Schechter, Skeletal muscle as an endogenous nitrate reservoir., *Nitric Oxide* 47 (2015) 10-16.
- [31] S.A. Omar, H. Fok, K.D. Tilgner, A. Nair, J. Hunt, B. Jiang, P. Taylor, P. Chowienczyk, A. Webb, Paradoxical Normoxia-Dependent Selective Actions of Inorganic Nitrite in Human Muscular Conduit Arteries, and Related Selective Actions on Central Blood Pressures., *Circulation* (2014)

- [32] J.O. Lundberg, Nitrate transport in salivary glands with implications for NO homeostasis., *Proc Natl Acad Sci U S A* 109 (2012) 13144-13145.
- [33] V. Kapil, A.J. Webb, A. Ahluwalia, Inorganic nitrate and the cardiovascular system, *Heart* 96 (2010) 1703-1709.
- [34] A.R. Coggan, J.L. Leibowitz, C.A. Spearie, A. Kadkhodayan, D.P. Thomas, S. Ramamurthy, K. Mahmood, S. Park, S. Waller, M. Farmer, L.R. Peterson, Acute Dietary Nitrate Intake Improves Muscle Contractile Function in Patients With Heart Failure: A Double-Blind, Placebo-Controlled, Randomized Trial., *Circ Heart Fail* 8 (2015) 914-920.
- [35] A.R. Coggan, J.L. Leibowitz, A. Kadkhodayan, D.P. Thomas, S. Ramamurthy, C.A. Spearie, S. Waller, M. Farmer, L.R. Peterson, Effect of acute dietary nitrate intake on maximal knee extensor speed and power in healthy men and women., *Nitric Oxide* 48 (2015) 16-21.
- [36] P. Zamani, D. Rawat, P. Shiva-Kumar, S. Geraci, R. Bhuvu, P. Konda, P.T. Doulias, H. Ischiropoulos, R.R. Townsend, K.B. Margulies, T.P. Cappola, D.C. Poole, J.A. Chirinos, Effect of inorganic nitrate on exercise capacity in heart failure with preserved ejection fraction., *Circulation* 131 (2015) 371-80; discussion 380.
- [37] T. Ashmore, B.O. Fernandez, C. Branco-Price, J.A. West, A.S. Cowburn, L.C. Heather, J.L. Griffin, R.S. Johnson, M. Feelisch, A.J. Murray, Dietary nitrate increases arginine availability and protects mitochondrial complex I and energetics in the hypoxic rat heart., *J Physiol* 592 (2014) 4715-4731.
- [38] A. Dejam, C.J. Hunter, C. Tremonti, R.M. Pluta, Y.Y. Hon, G. Grimes, K. Partovi, M.M. Pelletier, E.H. Oldfield, R.O. Cannon, A.N. Schechter, M.T. Gladwin, Nitrite infusion in humans and nonhuman primates: endocrine effects, pharmacokinetics, and tolerance formation., *Circulation* 116 (2007) 1821-1831.

- [39] A.R. Maher, S. Arif, M. Madhani, K. Abozguia, I. Ahmed, B.O. Fernandez, M. Feelisch, A.G. O'Sullivan, A. Christopoulos, A.L. Sverdlov, D. Ngo, R. Dautov, P.E. James, J.D. Horowitz, M.P. Frenneaux, Impact of chronic congestive heart failure on pharmacokinetics and vasomotor effects of infused nitrite., *Br J Pharmacol* 169 (2013) 659-670.
- [40] J.A. Chirinos, F. Londono-Hoyos, P. Zamani, M. Beraun, P. Haines, I. Vasim, S. Varakantam, T.S. Phan, T.P. Cappola, K.B. Margulies, R.R. Townsend, P. Segers, Effects of organic and inorganic nitrate on aortic and carotid haemodynamics in heart failure with preserved ejection fraction., *Eur J Heart Fail* 19 (2017) 1507-1515.
- [41] A.R. Coggan, S.R. Broadstreet, K. Mahmood, D. Mikhalkova, M. Madigan, I. Bole, S. Park, J.L. Leibowitz, A. Kadkhodayan, D.P. Thomas, D. Thies, L.R. Peterson, Dietary Nitrate Increases VO_2 peak and Performance but Does Not Alter Ventilation or Efficiency in Patients With Heart Failure With Reduced Ejection Fraction., *J Card Fail* 24 (2018) 65-73.
- [42] L.R. Peterson, K.B. Schechtman, G.A. Ewald, E.M. Geltman, L. de las Fuentes, T. Meyer, P. Krekeler, M.L. Moore, J.G. Rogers, Timing of cardiac transplantation in patients with heart failure receiving beta-adrenergic blockers., *J Heart Lung Transplant* 22 (2003) 1141-1148.
- [43] C.P. Kerley, J.O. O'Neill, V. Reddy Bijjam, C. Blaine, P.E. James, L. Cormican, Dietary nitrate increases exercise tolerance in patients with non-ischemic, dilated cardiomyopathy- a double-blind, randomized, placebo-controlled, crossover trial., *J Heart Lung Transplant* 35 (2016) 922-926.
- [44] D.M. Hirai, J.T. Zelt, J.H. Jones, L.G. Castanhas, R.F. Bentley, W. Earle, P. Staples, M.E. Tschakovsky, J. McCans, D.E. O'Donnell, J.A. Neder, Dietary nitrate supplementation and exercise tolerance in patients with heart failure with reduced ejection fraction., *Am J Physiol Regul Integr Comp Physiol* 312 (2017) R13-R22.

- [45] D. Stoyanovsky, T. Murphy, P.R. Anno, Y.M. Kim, G. Salama, Nitric oxide activates skeletal and cardiac ryanodine receptors., *Cell Calcium* 21 (1997) 19-29.
- [46] T.L. Dutka, J.P. Mollica, G.S. Posterino, G.D. Lamb, Modulation of contractile apparatus Ca^{2+} sensitivity and disruption of excitation-contraction coupling by S-nitrosoglutathione in rat muscle fibres., *J Physiol* 589 (2011) 2181-2196.
- [47] L. Nogueira, C. Figueiredo-Freitas, G. Casimiro-Lopes, M.H. Magdesian, J. Assreuy, M.M. Sorenson, Myosin is reversibly inhibited by S-nitrosylation., *Biochem J* 424 (2009) 221-231.
- [48] F.J. Larsen, E. Weitzberg, J.O. Lundberg, B. Ekblom, Effects of dietary nitrate on oxygen cost during exercise., *Acta Physiol (Oxf)* 191 (2007) 59-66.
- [49] F.J. Larsen, E. Weitzberg, J.O. Lundberg, B. Ekblom, Dietary nitrate reduces maximal oxygen consumption while maintaining work performance in maximal exercise., *Free Radic Biol Med* 48 (2010) 342-347.
- [50] F.J. Larsen, T.A. Schiffer, S. Borniquel, K. Sahlin, B. Ekblom, J.O. Lundberg, E. Weitzberg, Dietary inorganic nitrate improves mitochondrial efficiency in humans., *Cell Metab* 13 (2011) 149-159.
- [51] P.B. Massion, O. Feron, C. Dessy, J.L. Balligand, Nitric oxide and cardiac function: ten years after, and continuing., *Circ Res* 93 (2003) 388-398.
- [52] T. Münzel, A. Daiber, T. Gori, More answers to the still unresolved question of nitrate tolerance., *Eur Heart J* 34 (2013) 2666-2673.

Table 1. Comparison of the inorganic NO₃⁻ pathway with other sources of NO and/or cGMP

Other sources of NO/cGMP	Advantage of inorganic NO ₃ ⁻
L-arginine*	<ul style="list-style-type: none"> • Not dependent on NO synthase (NOS) • Functions well in acidic tissues • Functions well in ischemic tissues; does not require molecular oxygen
<i>Organic</i> , pharmacologic nitrates (e.g., nitroglycerin)**	<ul style="list-style-type: none"> • Does not cause tolerance • Does not increase reactive oxygen species (ROS) • May be less likely to cause hypotension† • May be less likely to cause flushing or headache† • Decreases left ventricular late systolic load • Does not cause cross-tolerance with nitroglycerin [39]
Phosphodiesterase 5 inhibitors (e.g., sildenafil)	<ul style="list-style-type: none"> • May be less likely to cause hypotension† • May be less likely to cause flushing or headache† • Less likely to cause retinal dysfunction and vision changes through inhibition of PDE6

*The substrate used for NOS-related production of NO.

**Not all pharmacologic nitrates are prone to these same disadvantages or to the same degree (see review by Munzel et al. [52]).

†These symptoms were not observed in our preliminary studies of HFrEF patients treated with 11.2 mmol of inorganic nitrate in the form of beetroot juice [34, 35, 41], and inorganic nitrate does not cause significant cerebrovascular dilation in HF with *preserved* ejection fraction [40].

Highlights

- Low nitric oxide (NO) likely contributes to exercise intolerance in heart failure.
- The inorganic nitrate pathway has distinct advantages over other sources of NO.
- Dietary inorganic nitrate, such as in beetroot juice, is chemically reduced to nitrite and then NO.
- Inorganic nitrate may improve both muscle power and aerobic exercise performance.

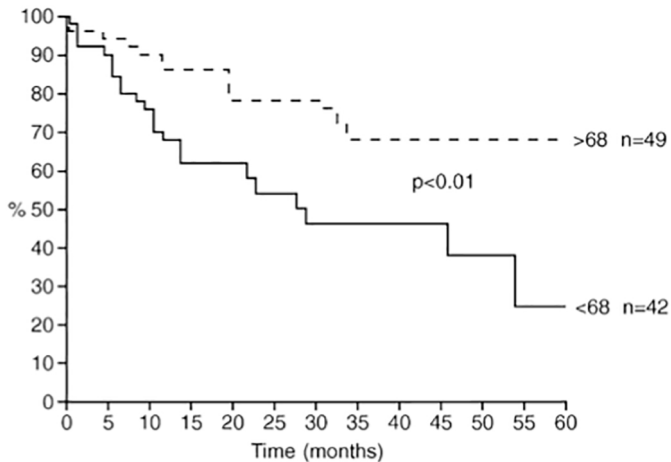


Figure 1

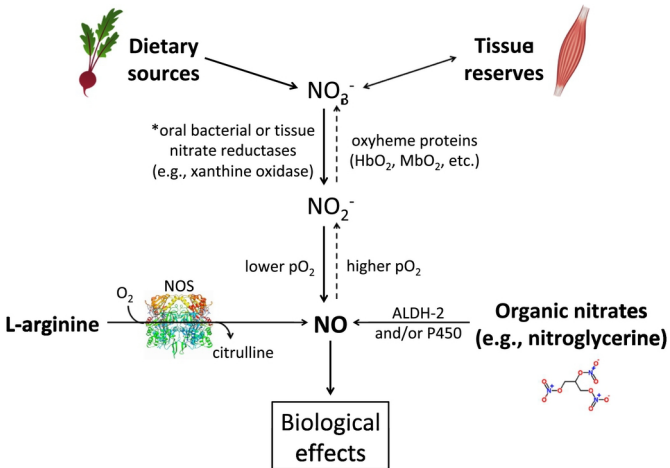


Figure 2